AWARD NUMBER: W81XWH-15-1-0124

TITLE: Novel Therapeutic Development of NF1-Associated Malignant Peripheral Nerve Sheath Tumor (MPNST)

PRINCIPAL INVESTIGATOR: Ping Chi, MD, PhD

CONTRACTING ORGANIZATION: Sloan Kettering Institute for Cancer Research New York, NY 10065

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13. SUPPLEMENTARY NOTES

14. ABSTRACT

NF1-assocaited MPNST accounts for ~50% of all MPNSTs and is one of the most difficult subtypes of soft tissue sarcoma to manage, given its poor sensitivity to conventional systemic therapy and radiotherapy and high likelihood to metastasize. Novel therapeutic development is imperative. Using comprehensive and integrative genomic approaches, we have identified recurrent inactivating genetic alterations of three central pathways: PRC2 (80%) core components (EED or SUZ12), CDKN2A (81%) and NF1 (72% of non-NF1-associated) in all MPNSTs, and the three components significantly co-occur, suggesting their critical and cooperative roles in MPNST pathogenesis. We demonstrated that MPNSTs with complete PRC2 loss showed complete loss of H3K27me3 and aberrant transcriptional activation of multiple PRC2-transcriptional targets. Here, we have further validated the high sensitivity and specificity of using H3K27me3 as a diagnostic biomarker for high-grade MPNST. The evaluation of using H3K27me3 as a prognostic biomarker in MPNST is ongoing. Therapeutically, we have found that the combination of MEK inhibitor and CDK4 inhibitor resulted in at least additive effects of growth suppression of human MPNST cell lines in vitro; We are currently evaluating newer generations of epigenetic therapeutics in PRC2-loss MPNST cells. These studies will be extended to in vivo xenograft studies and relevant genetically engineered MPNST mouse models.

15. SUBJECT TERMS

PRC2 loss, H3K27me3, CDK4/6 inhibitor, MEK inhibitor, MPNST, NF1-associated MPNST

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INTRODUCTION

Neurofibromatosis type 1 (NF1) is a common autosomal dominant disorder that affects approximately 1 in 3000 people. Although multiple defects can arise, malignant peripheral nerve sheath tumor (MPNST) represents the most common and deadly malignant tumors, leading to significant morbidity and mortality in NF1 patients. MPNSTs represent a group of highly aggressive soft tissue sarcomas that may occur in distinct clinical settings: NF1-associated (50%), sporadic (40%) or radiotherapy-associated (10%). Novel therapeutics is imperative. Using comprehensive and integrative genomic approaches, we have recently identified recurrent loss of function genetic alterations of three central pathways: PRC2 (80%) core components (EED or SUZ12), CDKN2A (81%) and NF1 (72% of non-NF1-associated) in all MPNSTs, and the three components significantly co-occur. These data suggest that, despite the clinical and histological heterogeneity, the NF1-associated, sporadic and RT-associated MPNSTs represent a relatively homogeneous group of disease at the molecular level. We also demonstrated that MPNSTs with complete PRC2 loss showed complete loss of H3K27me3 and aberrant transcriptional activation of multiple PRC2-repressed homeobox master regulators and their regulated developmental pathways. Introduction of the PRC2 component in a PRC2-deficient MPNST cell line restored H3K27me3 and decreased cell growth. The highly recurrent and specific inactivation of PRC2, NF1 and CDKN2A posits their critical and potentially cooperative roles in MPNST pathogenesis (Lee et al., Nat Genetics, in press). Based on these observations, we hypothesize that 1) H3K27me3 IHC may represent a novel prognostic and therapeutic biomarker in MPNST: 2) the frequent and concurrent CDKN2A loss. NF1 loss and PRC2 loss in all MPNSTs may create therapeutic opportunities for novel therapeutics development in all MPNSTs. Here, we focus on two specific aims: Aim1: Evaluate H3K27me3 by IHC as a prognostic biomarker in MPNSTs. Aim 2: Identify novel therapeutic strategies in MPNSTs using in vitro and in vivo models focusing on a) targeting CDKN2A loss and NF1 loss; b) targeting PRC2 loss.

KEYWORDS

PRC2 loss, H3K27me3, CDK4/6 inhibitor, MEK inhibitor, Malignant Peripheral Nerve Sheath Tumor (MPNST), Neurofibromatosis type I (NF1)-associated MPNST

ACCOMPLISHMENTS

Major goals of the project:

Aim1: Evaluate H3K27me3 by IHC as a prognostic biomarker in MPNST.

- 1) Clinical sample retrieval (Human FFPE samples) and pathology review
- 2) H3K27me3 IHC of MPNST FFPE samples
- 3) Clinical database evaluation
- 4) Statistical analysis
- 5) Local IRB/IACUC approval for proposed retrospective bio-specimen and animal studies.

Aim2: Identify novel therapeutic strategies in MPNSTs using *in vitro* and *in vivo* models focusing on a) targeting CDKN2A and NF1 loss and b) targeting PRC2 loss.

- 1) Targeting CDKN2A and NF1 loss using combination therapy of a CDK4/6 inhibitor and MEK inhibitor in human MPNST cells and MPNST xenograft mouse models.
- 2) Targeting PRC2 loss using P300/CBP inhibitors in human MPNST cells and MPNST xenogfrat mouse models.

Accomplished goals to date:

Aim1: Evaluate H3K27me3 by IHC as a prognostic biomarker in MPNST.

We have assembled a cohort of 68 FFPE samples from 58 MPNST patients, with 44 patients that have been previously described in our previous publication (Lee et al., Nature Genetics, 2014). We have performed H3K27me3 on these samples and compared it with other types of soft tissue sarcomas. We found that 69% of all MPNSTs have demonstrated loss of H3K27me3 mark, whereas 31% retained H3K27me3 expression. Among the NF1-assocaited high grade MPNSTs, 60% demonstrated loss of expression. In the sporadic and radiotherapy-associated MPNSTs, 95% and 91% of MPNSTs have demonstrated loss of H3K27me3,

respectively. 2/3 low-grade MPNST and all neurofibromas have demonstrated presence of H3K27me3. In addition, none of the 5 epithelioid MPNSTs, 53 cutaneous melanoms, 113 synovial sarcomas, 123 GISTs, 75 of well-differentiated and dedifferentiated liposarcomas, 6 Ossifying fibromyxoid tumor, 6 soft tissue myoepithelial carcinomas, and 63 myxofibrosarcomas have lost H3K27me3 IHC. These data demonstrates that the H3K27me3 IHC is a sensitive and specific biomarker for MPNST, particularly outside of NF1-clinical history, which represents the most challenging diagnostic scenarios. This collaborative work between the Chi lab and Dr. Antonescu's lab is published recently (Prieto-Granada CN, et al, American Journal of Pathology, 2016) (see below).

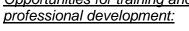
In the meantime, we have continued our efforts to work with Dr. Antonescu and Dr. Singer to prospectively identify more MPNST samples/patients with the goal to reach 100 samples for prognostic biomarker studies. We have obtained the IRB approval for the retrospective bio-specimen study entitled "Genomic and Biological Analyses of Malignant Peripheral Nerve Sheath Tumors (MPNST)" (IRB #16-052) on 2/11/2016. We have also obtained the revised IACUC approval to accommodate the in vivo xenograft studies for human MPNST cells for Aim 2 as well as annual renewal approved on 4/22/2016.

Aim2: Identify novel therapeutic strategies in MPNSTs using in vitro and in vivo models focusing on a) targeting CDKN2A and NF1 loss and b) targeting PRC2 loss.

We have evaluated the dual targeting of CDKN2A and NF1 loss using combination therapy of a CDK4/6 inhibitor (palbociclib) and a MEK inhibitor (trametinib) both have been FDAapproved for breast cancer and melanoma, respectively. We have assayed for growth suppressive effects of either single agent or in combination of these two drugs across different doses in three MPNST cell lines and observed additive growth suppressive effects in all three MPNST cell lines (Figure 1). These studies will move into xenograft studies in the next year.

We have also established MTA and obtained the newer generation of more potent P300/CBP inhibitors from AbbVie. We generated two isogenic MPNST cell lines that harbor NF1/CDKN2A loss or NF1/CDKN2A/PRC2 loss by CRISPRmediated deletion of EED or SUZ12. loss of either will inactivate PRC2. We tested the two P300/CBP compounds p300-1, p300-2 with inert compound control and observed a consistent but modest selective inhibitory activity of cell growth and survival of MPNST cells with PRC2 loss compared to PRC2-wild-type MPNST cells (data not shown). These studies will be extended to other MPNST cell lines with similar strategy for in vitro testing. If promising in multiple MPNST cell lines, we will move to in vivo xenograft studies.

Opportunities for training and professional development:



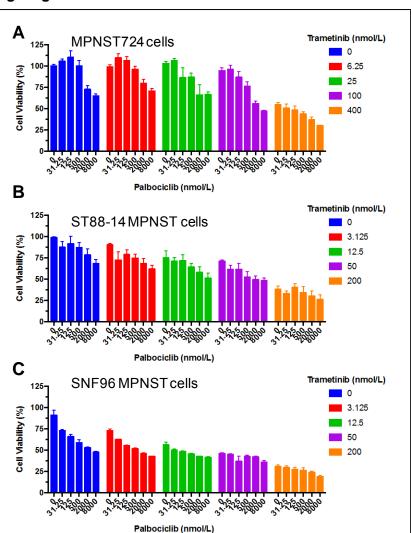


Figure 1. Cell viability assay with drug treatment at dose concentrations as indicated in various MPNST cell lines. All cells were treated for 7 days in triplicates and harvested and assayed for growth by CellTiter-Glo. A. MPNST724 (NF1-wild type, CDKN2A loss, and PRC2 wild-type). B. ST88-14 MPNST cells (NF1 loss, CDKN2A loss, PRC2 loss). C. SNF96 MPNST cells (NF1 loss, CDKN2A loss, PRC2 loss). Error bars: mean± SD.

Dissemination of results to communities of interest:

- 1) Invited talks at national/international meetings:
 - a) "The role of PRC2 components in the development of malignant peripheral nerve sheath tumor (MPSNT)", 11/5/2015, SARC-CTOS (Connective Tissue Oncology Society) Symposium, Salt Lake City, Utah
 - b) "PRC2 loss in malignant transformation of MPNST", 6/19/2016, 2016 NF conference, Austin, Texas
- 2) Publications:
 - a) Prieto-Granada CN, Wiesner T, Messina JL, Jungbluth AA, Chi P, Antonescu CR. Loss of H3K27me3 expression is a highly sensitive marker for sporadic and radiation-induced MPNST. Am J Surg Pathol 2016;40:479-89. PMID: 26645727. PMCID: PMC4882106.

Plans for next reporting period:

As described in the above, for Aim 1, we have continued our efforts to work with Dr. Antonescu and Dr. Singer to prospectively identify more MPNST samples/patients with the goal to reach 100 samples for prognostic biomarker studies.

For Aim 2, we will move the combination CDK 4/6 and MEK inhibitors to *in vivo* xenograft studies. We will expand the *in vitro* studies of P300/CBP inhibitors in more isogenic MPNST cell lines. If promising, we will move into *in vivo* xenograft studies in the next reporting perioid.

IMPACT

Impact on the development of the principal disciplines of the project/impact on other disciplines:

Based on our oncogenomics and clinical investigations, we have established H3K27me3 IHC as a sensitive and specific biomarker for diagnosis of MPNST especially in the non-NF1 setting. Our pathology group and many others have established the methodology and have routinely used it in current modern diagnoses.

The conceptual advances gained with our studies have benefited our hypothesis-driven investigations of novel therapeutic combinations. Their success will help facilitate newer generations of clinical investigations for MPNST.

Impact on technology transfer:

Nothing to report.

Impact on society beyond science and technology:

Our studies have established new diagnostic biomarkers that have been adapted by multiple pathology groups nation-wide.

CHANGES/PROBLEMS

Nothing to report.

PRODUCTS

Conference presentations-invited talks at national/international meetings:

- 1) "The role of PRC2 components in the development of malignant peripheral nerve sheath tumor (MPSNT)", 11/5/2015, SARC-CTOS (Connective Tissue Oncology Society) Symposium, Salt Lake City, Utah
- 2) "PRC2 loss in malignant transformation of MPNST", 6/19/2016, 2016 NF conference, Austin, Texas

Publications:

Prieto-Granada CN, Wiesner T, Messina JL, Jungbluth AA, **Chi P**, Antonescu CR. Loss of H3K27me3 expression is a highly sensitive marker for sporadic and radiation-induced MPNST. **Am J Surg Pathol** 2016;40:479-89. PMID: 26645727. PMCID: PMC4882106.

PARTICIPANTS & OTHER COLLABORATING ORGANIZATIONS

Individuals that have worked on the project

Principal Investigator: Ping Chi, MD, PhD (HOPP and Department of Medicine, MSKCC) (1.2 cal mo) (No change)

Collaborator: Cristina R. Antonescu, MD (Department of Pathology, MSKCC) (0.6 cal mo) (No change)

Collaborator: Sam Singer, MD (Department of Surgery, MSKCC) (0.6 cal mo) (No change)

Collaborator: Li-Xuan Qin, PhD (Biostatistics, MSKCC) (0.6 cal mo) (No change)

Leili Ran, PhD, was a graduate student and has recently graduated with PhD and transitioned into a postdoc position and continued the project. (12.0 cal mo)

Amish Patel, PhD, a new postdoc, has been working on the P300/CBP inhibitor studies. (2.4 cal mo)

Miguel A. Miranda-Roman, B.S. a graduate student, has been working on the *in vitro* studies of CDK4/6 and MEK inhibitors. (1.2 cal mo)

Devan Murphy, a research assistant, has moved on to graduate school and her role has been replaced by Jessica Sher, a Research assistant and continued her responsibilities. (6.0 cal mo)

Changes in Active Support of the PI

COMPLETED:

5 K08 CA151660-4 (PI: Chi) 07/01/11-06/30/16 7.8 calendar months

NIH/NCI \$157,027

Title of Project: Elucidating the roles of ETV1 in the pathogenesis of GIST.

The goal of the project is to investigate the genetic and epigenetic regulation of ETV1-dependent oncogenic transcriptome in GIST oncogenesis and to establish the oncogenic role of ETV1 involved in the initiation and/or maintenance of GIST *in vivo*.

Specific Aims:

Aim 1: Define ETV1 cistrome and integrate with transcriptional output in the ICC-GIST lineage.

Aim 2: Dissect the functional interplay between KIT signaling and ETV1 in GIST.

Aim 3: Investigate the role of ETV1 for initiation and maintenance of GIST in mouse models.

Agency Contact: Esther Young (Grants Management Specialist)

Email: youngel@mail.nih.gov, Phone: (240) 276-6325

5 P50 CA140146-05 (PI: Singer) 11/01/13-10/31/15 0.60 calendar months

NIH/NCI \$211.904

SPORE in Soft Tissue Sarcoma

Project 1 Title: Molecular Mechanisms of KIT signaling and Imatinib Resistance in GIST

The goal is to perform biochemistry and oncogenomics, as well as using imatinib-resistant GIST murine models to dissect the resistance mechanisms in imatinib-resistant GIST.

Agency Contact: Renee Carruthers (Grants Management Specialist)

Email: carruthersr@mail.nih.gov, Phone: 301-631-3018, Fax: 301-451-5391

Kimmel Scholar Award (Pl: Chi) 07/01/12-06/30/15* 0.0 calendar months

Sidney Kimmel Foundation for Cancer Research \$86,957

Understanding and targeting the oncogenic lineage-specific transcription factor- ETV1-in Gastrointestinal Stromal Tumor (GIST)

The goal of this project is to characterize the genetic and epigenetic mechanisms by ETV1 that define the optimal cellular context for mutant KIT-driven oncogenesis, and to dissect the molecular mechanisms of ETV1 protein stability regulation.

Specific Aims:

- 1. How is the lineage-specific and ETV1-dependent oncogenic transcriptome regulated in GIST?
- 2. How is ETV1 protein stability regulated by KIT/MAP kinase signaling pathway and can we target ETV1 protein stability in imatinib-resistant GIST?
- 3. Can we target ETV1 transcriptional activity in GIST and other ETV1-dependent malignancies? Agency Contact: Gary I. Cohen, MD, FACP

Email: gcohen@gbmc.org, Phone: (443) 849-3729

ACTIVE:

5 DP2 CA174499-04 (PI: Chi) 09/13/12-8/31/17 3.00 calendar months

NCI \$300,000

An integrative approach to target lineage-specific oncogenic transcription factors

This proposal describes a novel integrative approach that combines a high throughput RNAi-based screen with high complexity gene-signature based readout to systematically discover druggable modifiers of oncogenic transcription factors.

Specific Aims:

1. ETV1 is oncogenic in prostate cancer, GIST, and melanoma

2. ETV1 regulates distinct lineage specific genes in prostate cancer and GIST

3. Combined RNAi and high complexity GE-HTS for specific targeting of ETS-dependent transcriptomes Agency Contact: Andrea Bell (Grants Management Specialist)

Email: bellan@mail.nih.gov Phone: 301-496-3276, Fax: 301-496-8601

W81XWH-15-1-0124 (PI: Chi) 07/15/15-07/14/18 1.20 calendar months

Congressionally Directed Medical Research \$133,280

Programs

Novel therapeutic development of NF1-assocaited malignant peripheral nerve sheath tumor (MPNST)

Aim 1: Evaluate H3K27me3 by IHC as a prognostic biomarker in MPNSTs. Aim 2: Identify novel therapeutic strategies in MPNSTs using *in vitro* and *in vivo* models focusing on a) targeting CDKN2A loss and NF1 loss; b) targeting PRC2 loss.

Agency Contact: Wendy A Baker

Email: wendy.a.baker.civ@mail.mil, Phone: 301-619-2034

SPECIAL REPORTING REQUIREMENTS:

None.

APPENDICES:

- 1) Approval of Protocol # 16-052, "Genomic and Biological Analyses of Malignant Peripheral Nerve Sheath Tumors (MPNST)"
- 2) Journal article:

Prieto-Granada CN, Wiesner T, Messina JL, Jungbluth AA, **Chi P**, Antonescu CR. Loss of H3K27me3 expression is a highly sensitive marker for sporadic and radiation-induced MPNST. **Am J Surg Pathol** 2016;40:479-89. PMID: 26645727. PMCID: PMC4882106.



Institutional Review Board/Privacy Board

TO: Ping Chi, MD, PhD

Department of Medicine/Sarcoma Medical Oncology Service

Roger S. Welson

FROM: Roger S Wilson, MD

Chairman, Institutional Review Board/Privacy Board-A

DATE: 02/11/2016

RE: Protocol # 16-052

Your protocol entitled "Genomic and Biological Analyses of Malignant Peripheral Nerve Sheath Tumors (MPNST)", was re-reviewed on 02/11/2016 by the Institutional Review Board/Privacy Board-A and was approved by expedited review process as per 45 CFR 46.110 and 21 CFR 56.110 for 12 months.

Your request to waive the requirement to obtain informed consent and a research authorization for the sample analysis outlined in your biospecimen protocol has been granted as per 45 CFR 46.116(d) and 45 CFR 164.512(i)(2)(ii). Participant informed consent must be obtained under the approved specimen collection studies outlined in section 6.0 of this protocol.

Type of Biomaterial	Disease Name	Disease Site	Sample required	Amount required for each case	Number of cases required	Source
Primary	Other	NO SITE REQMT	Frozen	50	40	Banked Specimens (MSKCC)
Companion Normal	Other	NO SITE REQMT	Frozen	50	40	Banked Specimens (MSKCC)
Metastatic	Other	NO SITE REQMT	Frozen	50	40	Banked Specimens (Collaborator)
Primary	Other	NO SITE REQMT	Paraffin Block	25	200	Banked Specimens (MSKCC)
Metastatic	Other	NO SITE REQMT	Paraffin Block	25	200	Banked Specimens (MSKCC)

IRB-A Determination of Risk

Protocol: Low Children: N/A Device: N/A

Genetic Research: Yes; GWAS: No TCGA: No



HHS Public Access

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Am J Surg Pathol. 2016 April; 40(4): 479–489. doi:10.1097/PAS.000000000000564.

Loss of H3K27me3 Expression Is a Highly Sensitive Marker for Sporadic and Radiation-induced MPNST

Carlos N. Prieto-Granada, MD*,†, Thomas Wiesner, PhD‡, Jane L. Messina, MD†, Achim A. Jungbluth, MD*, Ping Chi, MD, PhD‡,§,∥, and Cristina R. Antonescu, MD*

*Department of Pathology, Memorial Sloan Kettering Cancer Center

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Department of Medicine, Weill Cornell Medical College, New York, NY

†Moffitt Cancer Center (MCC), Tampa, FL

Abstract

Most malignant peripheral nerve sheath tumors (MPNSTs) exhibit combined inactivation of NF1, CDKN2A, and polycomb repressive complex 2 component genes (Embryonic Ectoderm Development [EED] and Suppressor of Zeste 12 [SUZ12]). Mutations in EED and SUZ12 induce loss of trimethylation at lysine 27 of histone 3 (H3K27me3), with subsequent aberrant transcriptional activation of polycomb repressive complex 2-repressed homeobox master regulators. These findings prompted us to investigate the performance of an anti-H3K27me3 monoclonal antibody clone C36B11 as an immunohistochemical marker for MPNSTs. We assessed the C36B11 reactivity pattern in a pathologically and genetically well-characterized cohort of 68 MPNSTs, spanning various clinical presentations, such as type 1 neurofibromatosis (NF1), radiotherapy, and sporadic MPNSTs. We found that 69% (n=47) of all MPNSTs demonstrated loss of H3K27me3 expression, with 42 (61%) showing complete loss and 5 (7%) showing partial loss, whereas 31% (n = 21) retained H3K27me3 expression. Among the NF1related high-grade MPNSTs, 60% demonstrated loss of expression. In contrast, the majority of both sporadic (95%) and radiotherapy-related (91%) MPNSTs showed loss of H3K27me3 expression. Two of the 3 low-grade MPNSTs and all neurofibromas showed retained expression. Furthermore, all 5 epithelioid MPNSTs retained H3K27me3 labeling. The specificity of H3K27me3 loss as a marker for MPNSTs was studied by testing a large spectrum of lesions included in MPNST differential diagnosis, such as spindle/desmoplastic melanomas, synovial sarcomas, myoepithelial tumors, and other mesenchymal neoplasms, all of which retained expression of H3K27me3. We conclude that immunohistochemical analysis of H3K27me3 has

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Conflicts of Interest and Source of Funding: The authors have disclosed that they have no significant relationships with, or financial interest in, any commercial companies pertaining to this article.

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good sensitivity and robust specificity for the diagnosis of MPNST, particularly outside of NF1 clinical history, which represents the most challenging diagnostic setting.

Keywords

malignant peripheral nerve sheath tumor; neurofibromatosis 1; polycomb repressor complex 2; EED; SUZ12; H3K27me3

Malignant peripheral nerve sheath tumors (MPNSTs) are rare neoplasms occurring mostly in adults and representing approximately 4% of all sarcomas. About 50% of MPNSTs occur in patients afflicted by type 1 neurofibromatosis (NF1), whereas about 10% of cases arise secondary to prior radiotherapy (RT).^{2,3} The remaining 40% of cases occur without known predisposition and include both conventional/spindle and epithelioid cell variants. A 2-tiered grading system with low-grade (LG) and high-grade (HG) MPNSTs is used, with the LG end being characterized by neurofibroma (NF)-like tumors composed of a cellular proliferation of bland, back-to-back spindle cells that appear to be in a continuum with the so-called "atypical/cellular neurofibroma." 4,5 Histologically, HG MPNSTs are characterized by an undifferentiated phenotype with limited, if any, evidence of schwannian differentiation at immunohistochemical (IHC) or ultrastructural levels. The majority of MPNSTs demonstrate a characteristic pattern of intersecting fascicles of relatively monotonous spindle cells with hyperchromatic nuclei, very high mitotic counts, and geographic areas of necrosis. This monomorphic spindle cell appearance is shared with other soft tissue sarcomas, chiefly monophasic synovial sarcomas (SSs) and adult-type fibrosarcomas among others. ^{4,6} Rare morphologic variants include the pleomorphic type and MPNSTs with perineuriomatous differentiation.^{4,7} MPNSTs display limited expression of neural crest markers, such as S100 protein and Sox10, with only 30% to 39% and 49% to 67% of cases, respectively, showing positivity for these markers according to recent studies.^{8–10} In contrast, epithelioid MPNSTs consistently show diffuse S100 protein positivity and loss of INI1/SMARCB1 expression. Thus, accurate diagnosis of non-epithelioid MPNSTs outside the NF1 or radiation clinical history remains challenging because of lack of specific IHC and molecular markers.

Recently, our group and others demonstrated loss-of-function somatic alterations in different components of the polycomb repressive complex 2 (PRC2) in the majority of MPNST cases. ^{11–13} These highly recurrent and specific inactivations of PRC2 components cooccurred with somatic alterations of *CDKN2A* and *NF1*. MPNSTs with PRC2 inactivation through *EED* or *SUZ12* alterations showed consistent and complete loss of trimethylation at lysine 27 of histone H3 (H3K27me3) on IHC analysis. H3K27me3 represents an important intermediary of the PRC2 pathway of chromatin regulation. ^{14,15} Although positive and negative H3K27me3 immunostaining was highly concordant with the genetic status of wild-type (WT) PRC2 components and homozygous loss of PRC2 components, respectively, ¹¹ heterozygous loss of PRC2 components was not predictive of H3K27me3 immunostaining. In fact, analysis of H3K27me3 expression at a protein level appeared to be more accurate than DNA sequencing, as reflected by RNA sequencing and transcriptional clustering. On

the basis of these findings, we sought to investigate the specificity and sensitivity of a new monoclonal H3K27me3 antibody as an ancillary diagnostic marker in MPNSTs.

Materials and Methods

The diagnosis of MPNST was reviewed and confirmed on the basis of a constellation of findings including typical morphologic patterns, focal/patchy S100 protein/Sox10 reactivity (defined as <20% of tumor cells) or nonspecific immunoprofile, presence of a preexistent benign peripheral nerve sheath tumor, and clinical history of NF1 or prior radiation. The typical morphologic picture defining a conventional HG MPNST is represented by a fascicular, monomorphic spindle cell proliferation with a "marbled" low-power appearance and areas of geographic necrosis. Tumors with these characteristics were classified as having "classic" morphology (Figs. 1A, B). The diagnosis of LG MPNST was defined by the above-described morphologic features. Finally, examples exhibiting histomorphologic features that deviated from these classic LG and HG patterns were classified under MPNST with variant morphology, which mainly included tumors exhibiting pleomorphism (focal or diffuse) and a rare tumor showing small cell features, all these phenotypes being previously described in the literature.^{5,7} To accept a diagnosis of MPNST in the setting of nuclear pleomorphism other strict criteria were required, such as NF1 history, origin from a nerve or benign peripheral nerve sheath tumor, and/or focal S100 protein positivity (<20%) in the absence of other markers. The epithelioid MPNST cases displayed distinctive morphologic features with solid or nested growth of purely epithelioid cells, showing diffuse and strong S100 protein positivity and INI1/SMARCB1 loss. An effort was made to include cases with a complete IHC workup available for review, as well as cases with additional material for IHC analysis. The study was approved by the Institutional Review Board of both institutions.

MPNST Patients and Tumor Samples

Our cohort consisted of a total of 68 samples from 58 MPNST patients, with 44 patients being previously included in the study by Lee et al. 11 The selection of MPNST cases in that study was based on typical morphology, matching immunoprofile, and clinical findings, double-blinded from the molecular results. We centered our investigation on the prior MPNST study group to correlate the IHC findings of a new anti-H3K27me3 monoclonal antibody with the specific genetic alterations of PRC2 components and investigate the concordance with a previous IHC analysis using a H3K27me3 polyclonal antibody¹¹ (see antibody characteristics). The study was further expanded with additional MPNST cases, as well as other entities frequently included in the differential diagnosis. There were 3 groups of samples included: from patients with NF1, from patients with a history of RT to the tumor site, and from patients without any prior history (sporadic). An effort was made to include matched samples from the same patient exhibiting different morphologic features and/or from different time points. Thus, when available we included NF-MPNST pairs from the same resection, including examples of so-called atypical NFs (when possible in near proximity), areas of different morphology (classic MPNST, pleomorphic MPNST, and tumors with divergent differentiation component), matched primary-metastases, as well as multiple primary MPNSTs arising in the same patient.

There were 32 male (55%) and 26 female (45%) patients, with an overall mean age at diagnosis of 42 years (range, 2 to 76y). The average age at diagnosis varied depending on the clinical setting of MPNST. The NF1-related MPNST patients presented at a younger age, with a mean of 37 years (range, 2 to 59y), whereas the patients with sporadic MPNST presented at a mean age of 48 years (range, 28 to 76y). In RT-related MPNST patients, tumors presented at an older age, with a mean of 51 years (range, 32 to 75y). Five epithelioid MPNST patients were included, with a mean age of 37 years (range, 26 to 59y). There were 27 NF1 MPNST patients (47%), 15 patients with sporadic MPNST (26%) and 11 patients with RT-related MPNSTs (19%). The 5 patients with epithelioid MPNSTs (9%) were grouped separately from those with conventional spindle cell sporadic MPNSTs.

Control Groups of Tumors Included in the Differential Diagnosis of MPNST

To test the specificity of this marker, we analyzed different entities that are often considered in the differential diagnosis of MPNST, as well as miscellaneous tumors as controls. This process was carried out by applying a monoclonal anti-H3K27me3 antibody to tissue microarray (TMA) slides as well as to whole-tissue sections (WTS) with examples of such entities. The majority of sarcoma tumor samples were obtained from the Memorial Sloan Kettering Cancer Center Pathology Department, whereas the melanoma samples were derived from Moffitt Cancer Center Pathology Department. A TMA containing a total of 53 cutaneous melanomas from the same number of patients was tested, including 51 primaries/ recurrent lesions and 2 metastatic tumors to the lung. The melanoma cases were subdivided into 37 pure desmoplastic melanoma (DM) variants, 11 mixed DM with spindle and epithelioid components, and 5 spindle cell melanomas (SCM). All but 8 (15%) melanoma tumors (1 SCM, 2 pure DM, and 5 mixed DM/SCM) showed evidence of a melanoma in situ component. Immunoprofile data were available in all cases with the exception of 4 pure DM cases (7%), all of which showed melanoma in situ as well as classic pure DM features, both histopathologically (hypocellular proliferation of bland spindle cells in a sclerotic/myxoid stroma) and clinically (lack of lymph node metastases). All 48 tumors with available IHC exhibited diffuse and strong S100 positivity, and 24 of these tumors also showed the classic loss of Melan A and HMB45 staining. In addition, a TMA containing a large number (113) of SSs, including monophasic, biphasic, and poorly differentiated samples, was also studied. Also included were 6 soft tissue myoepithelial tumors (WTSs), 5 of them carrying the characteristic EWSR1 or FUS gene rearrangements. We investigated a TMA of 123 gastrointestinal stromal tumors (GISTs) from 113 patients, as well as WTS from KIT-mutant adult GIST, SDHB-deficient WT pediatric and adult GIST (13 cases), and KIT-negative dedifferentiated GIST (1 case). Finally, TMAs containing 75 liposarcomas (31 well differentiated and 44 dedifferentiated) and 63 myxofibrosarcomas (MFSs) (both LG and HG), as well as 6 WTS of ossifying fibromyxoid tumors (OFMTs), were tested.

H3K27me3 IHC

IHC for H3K27me3 was performed utilizing rabbit monoclonal antibody, clone C36B11 (1:200 dilution; Cell Signaling Technology, Danvers, MA). All immunostaining studies were performed on a Leica Bond-RX and Bond-3 automated stainer platform (Leica, Buffalo Grove, IL) employing a polymeric secondary detection system (Refine; Leica), using diaminobenzidine as a chromogen. The immunostaining protocol was tested and optimized

using samples from our pervious MPNST study using a rabbit polyclonal anti-HeK27me3 antibody (07-449; EMD Millipore, Billerica, MA). C36B11 was tested on TMA and WTS slides. Evaluation of the C36B11 IHC signal was recorded as positive/partially negative/ negative according to the status of the nuclear signal, and the proportion of lesional cells showing lack of nuclear staining with the marker was also semi-quantitatively recorded as complete (95%) or partial (<95%) loss of staining. The presence of positive internal controls (endothelial cells, stromal cells, and/or inflammatory cells) was strictly required to assess the IHC signal; samples not reaching this standard were excluded. The unpaired *t* test was used to assess for correlation between the different morphologic variables and H3K27me3 status.

Results

Clinicopathologic Features

A total of 68 MPNST samples from 58 patients were included in the study. Most samples (84%) originated from primary or recurrent lesions, whereas in 16% the available material was obtained from the metastatic lesions. The primary tumors were located in the limbs in 22 (37%) cases (upper limbs, 10; lower limbs, 12), in the trunk in 15 (26%) cases (11 paravertebral and 4 chest/mediastinum), in the abdominal region in 15 (26%) cases (retroperitoneum, 8; pelvis, 5; abdominal wall, 2), and in the head and neck in 5 (11%) cases. The majority of metastatic samples were located in the lung (9 cases), with 1 each in the mediastinum and adrenal gland. The primary lesions from which these metastases originated occurred in the lower limb (5 tumors), upper limb (2 tumors), and in the retroperitoneum and sacrum (1 each). Eighteen samples (26%) were status post prior chemoradiation therapy. In addition, 11 NFs were included, 8 being away from and 3 adjacent to the MPNST (evaluated in the same tissue section).

Of the 63 conventional (nonepithelioid) MPNSTs, 60 tumors (95%) showed an HG MPNST morphology, with the majority (72%) exhibiting a classic phenotype (Figs. 1A, B, Fig. 3C). Three cases (5%) showed LG MPNST features; all of them were NF1 related, with 1 tumor being associated with an atypical NF (MPNST13.1, 13.2). In the HG MPNST group (n = 60) there was a subset of 17 tumors (28%) exhibiting morphologic deviation from the classic features: 6 MPNSTs with pleomorphic features (4 NF1, 1 sporadic, and 1 RT) (Figs. 1D, E, Fig. 3E), 11 mostly classic MPNST but with focal pleomorphism(4NF1,4sporadic, and3RT) (Fig. 3A), and 1 tumor with focal pleomorphism and small cell features (sporadic, also displaying heterologous elements, Fig. 4D) Among these MPNSTs with variable pleomorphism, 4 (2 NF1 patients, 1 sporadic, and 1 RT) showed considerable morphologic intertumoral and intratumoral heterogeneity, with a classic HG MPNST component being invariably present in all cases. A total of 9 cases (13%) exhibited features of divergent differentiation (Fig. 4), 3 of them being NF1 related, 2 being RT induced, and the remaining 4 being sporadic. The most common heterologous component was rhabdomyosarcomatous (5 cases) (Figs. 4A, B), followed by glandular (3 cases) (Fig. 4B), chondrosarcomatous (2 cases), osteosarcomatous (1 case), and leiomyosarcomatous (1 case) (Fig. 4E). In 4 cases, the divergent differentiation consisted of 2 heterologous components (Figs. 4A, B). All 5 epithelioid MPNSTs exhibited the classic morphologic features of solid or nests of medium-

sized to large-sized epithelioid cells bearing eosinophilic to amphophilic cytoplasm and vesicular nuclei with prominent nucleoli, in a background of a fibrotic and focally myxoid stroma (Figs. 2D, E).

The Majority of Sporadic and RT-related MPNSTs Showed Loss of H3K27me3 Expression

IHC evaluation with the anti-H3K27me3 monoclonal antibody revealed that 69% (n= 47) of MPNST cases demonstrated either complete (n=42, 61%) (Figs. 1C, F, Fig. 3B, D) or partial (n = 5, 7%) (Fig. 2C) loss of staining, whereas 31% (n = 21) of cases retained diffuse immunoreactivity (Fig. 3F). However, considerable differences were observed in terms of C36B11 immunostaining status among the different MPNST groups. When considering only the NF1-related HG MPNSTs (n=30), 18 cases (60%) demonstrated loss of H3K27me3 expression (14 tumors showing complete loss and 4 lesions showing partial loss), whereas H3K27me3 expression was retained in the remaining 12 (40%) NF1-related HG MPNSTs. Among the 3 LG MPNSTs only 1 tumor showed loss of H3K27me3 expression (MPNST02). All 11 NFs, either distant or adjacent to the MPNSTs, retained C36B11 nuclear labeling (Fig. 1C, right aspect of picture). In contrast, the majority of both sporadic and RT-related MPNSTs showed loss of H3K27me3 expression, in 95% (17/18) and 91% (10/11) of cases, respectively. Only 1 of the sporadic MPNST showed partial loss of H3K27me3 (MPNST41). All 5 epithelioid MPNSTs retained H3K27me3 expression (Fig. 2F). See Table 1 for a summary of results.

There was no correlation (P= 0.25) between the presence of pleomorphic histology (n = 17) and the anti-H3K27me3 IHC status, with 12 (70%) tumors showing loss of H3K27me3 (11 total and 1 partial) and 5 retaining expression. When considering the different MPNST subgroups, differences can be noted with half of these cases occurring in the NF1 setting (n = 4) showing loss of staining (3 complete loss and 1 partial loss), whereas 5/6 sporadic and all 3 RT tumors exhibited complete loss of H3K27me3 expression. Among the 9 MPNSTs with divergent differentiation all except 1 (MPNST27, NF1-related) showed loss of H3K27me3 expression in the homologous component. H3K27me3 expression in the heterologous component was concordant with the homologous component in 2 of 4 cases evaluated, 2 showing loss, whereas 2 retained H3K27me3 expression (glandular and leiomyosarcomatous) (Fig. 4C, arrowheads and Fig. 4F, leiomyosarcoma "LMS"), suggesting intratumoral heterogeneity of H3K27me3 staining and perhaps of the mutational status of PRC2 gene components.

More than half (58%) of our conventional HG MPNST cohort exhibited at least focal positivity for S100 protein, whereas the remaining (42%) was completely S100 protein negative. No relationship was detected (P= 0.33) between the presence/absence of S100 immunostaining and the H3K27me3 IHC/PRC2 mutational status. The 2 retroperitoneal sporadic tumors (MPNST30 and 31) were negative for MDM2 and CDK4 immunoexpression. All 5 epithelioid MPNSTs were diffusely S100 protein positive, and 2 of the tumors tested with INI1/SMARCB1 (MPNST54, 58) also showed loss of expression of this marker.

Loss of H3K27me3 Expression Correlates With Genetic Alterations of PRC2 Components

The great majority of the MPNST cases with available molecular data (93%, 50/54) showed a good correlation between the H3K27me3 expression pattern (C36B11 immunostaining) and the PRC2 component genotype (EED and/or SUZ12). Cases with complete loss of C36B11 immunoreactivity correlated with either homozygous gene losses or point, frameshift, or splice mutations and structural variations, occurring alone or in combination with heterozygous losses. The "mosaic" pattern of partial loss of H3K27me3 expression detected in 4 NF1-related MPNSTs was associated with a SUZ12 heterozygous deletion and a WT EED in 3/4 cases, whereas a WT SUZ12 and EED heterozygous loss was found in the remaining case. No molecular data were available for the sporadic MPNST case showing partial loss of staining. Although H3K27me3-retained expression was seen in the majority of the WT/WT EED/SUZ12 genotype, there were 4 NF1-related tumors (MPNST16.1, 16.2, 17, and 18.1) in which discordant IHC/molecular results were found. Whereas the first 2 (16.1 and 16.2) showed lost H3K27me3 expression with a WT/WT EED/SUZ12 status, the other 2 exhibited diffuse H3K27me3 immunoreactivity in the presence of a SUZ12 heterozygous gene loss. A concordant pattern of WT/WT EED/SUZ12 profile and retained H3K27me3 expression was seen in all NF examples (n = 8) tested. For a complete list of the clinicopathologic findings and the mutational status of EED and SUZ12 please see Supplementary Tables 1 to 4 (Supplemental Digital Content 1, http://links.lww.com/PAS/ A321).

When comparing immunoreactivity of the monoclonal antibody C36B11 with that of the polyclonal antibody used in our previous study, only 2/52 (4%) MPNST tumors and NFs were found to demonstrate discordant immunoreactivity 11 (immunopositive with polyclonal antibody, immunonegative with C36B11 monoclonal antibody). Although there was an excellent correlation of the immunostaining of the 2 antibodies, the signal to noise ratio was superior with the monoclonal H3K27me3 antibody, which showed a crisper appearance with less background staining. This feature appears particularly important in the context of evaluating an IHC stain for loss of signal.

Intertumoral Heterogeneity of PRC2 Component Aberrations and H3K27me3 Expression Was Observed in Rare NF1-related MPNST Cases

Two examples in this category require a more detailed discussion. The first NF1-related MPNST exhibited different H3K27me3 expression patterns as well as different PRC2 component mutation profiles in the primary tumor (MPNST11.1) compared with the metastasis (MPNST11.2). Whereas the primary tumor exhibited complete loss of H3K27me3 expression due to an *EED* frameshift mutation and a *SUZ12* heterozygous deletion, the metastatic deposit showed a WT *EED* and *SUZ12* heterozygous deletion associated with partial loss of H3K27me3 expression.

The second NF1-related MPNST (MPNST15) developed 3 distinct soft tissue HG MPNSTs, with tissue available for IHC analysis of all lesions and molecular data available for analysis of 2/3 lesions. Morphologically, 1 tumor showed a classic HG phenotype with focal pleomorphism (MPNST15.1, retroperitoneum) (Fig. 3A), the second exhibited a classic HG phenotype (MPNST15.3, abdominal wall) (Fig. 3D), and the third primary was a

pleomorphic MPNST (MPNST15.4, thigh) (Fig. 3E). Both the MPNST with focal pleomorphism (MPNST15.1) and the classic HG MPNST (MPNST15.3) showed complete loss of H3K27me3 (Figs. 3B, D), with MPNST15.1 harboring an *SUZ12* frameshift mutation and heterozygous loss. The predominantly pleomorphic MPNST (MPNST15.4) revealed retained H3K27me3 expression (Fig. 3F), correlated with a WT/WT *EED/SUZ12*.

H3K27me3-retained Expression on IHC Study in All Look-alike Spindle Cell Neoplasms Tested and Other Miscellaneous Tumors

All tumors included as controls (see the Materials and methods section) exhibited positivity with the monoclonal antibody C36B11 to H3K27me3 (Fig. 5). A peculiar nucleolar pattern of signal accentuation was observed in some cases, particularly of SS (Fig. 5C) of uncertain significance. For a summarized display of these data, please refer Table 2.

Discussion

The diagnosis of MPNSTs remains challenging and somewhat arbitrary because of its nonspecific morphologic features and lack of pathognomonic ancillary IHC and molecular diagnostic markers. As a consequence, a high interobserver variability exists, even among experts, particularly when MPNST is encountered outside the known predisposing conditions (NF1, RT). Historically, the finding of either focal S100 positivity in 50% to 90% of cases ^{16–19} or frequent allelic losses (20% to 50%) of *NF1* in a background of complex karyotypes ²⁰ was considered consistent but relatively nonspecific features in these tumors. A more definitive diagnosis of MPNST was traditionally rendered in the context of particular clinical settings, such as in individuals affected by NF1 and tumors arising in a previously radiated area. When confronted with cases outside this clinical background (ie, in sporadic cases), the diagnosis of MPNST was often made on the basis of matching morphologic features, patchy S100 protein immunostaining, possible involvement of a nerve structure, and quite frequently as a diagnosis of exclusion from other spindle cell neoplasms.

A recent study from our group¹¹ characterized novel molecular aberrations in MPNSTs that affected the *EED* (11q14.2-q22.3) and *SUZ12* (17q11.2, formerly known as *JJAZ1*) genes, both of which code for homonymous proteins that form part of the PRC2. These aberrations were present in the majority (80%) of MPNSTs arising in a variety of clinical settings, such as NF1, RT, and sporadic scenarios, being in most cases mutually exclusive. These results were also independently confirmed and replicated by others. ^{13,21} The *EED* and *SUZ12* gene products, along with EZH1 and EZH2, constitute the core of the PRC2 complex and are chiefly responsible for the dimethylation and trimethylation of lysine 27 of histone H3 (H3K27me2 and H3K27me3), important epigenetic mediators of PRC2 function. ^{14,15} PRC2 is heavily involved in chromatin compaction, heterochromatin formation, chromosome X inactivation, and polycomb-mediated gene silencing. ^{22,23} Interestingly, *SUZ12* somatic recombinations have been described in patients with NF1 mosaicism, ^{24,25} which in part may explain the mosaicism encountered in our NF1-related cohort.

Our initial study showed that PRC2 inactivation through mutations in *EED* and *SUZ12* induced loss of H3K27me3 IHC expression using a polyclonal anti-body. On the basis of these findings, we sought to investigate the performance of a new monoclonal H3K27me3

antibody as an ancillary diagnostic marker in MPNST. Thus, we expanded the initial MPNST cohort as well as investigated a large spectrum of lesions typically included in the differential diagnosis of MPNST. Our results showed loss of H3K27me3 expression in 69% of all MPNSTs (NF1, RT related, sporadic, and epithelioid). In isolation, this result appears to have an equivalent diagnostic value to S100 protein or Sox10 IHC staining. However, when considering the different MPNST clinical subsets, loss of H3K27me3 expression emerges as a highly sensitive marker, being seen in most RT-related and sporadic MPNSTs. This finding reinforces the pathologic and molecular identity of MPNSTs even in lessdefined diagnostic circumstances, such as in non-NF1 patients. Although it remains uncertain why NF1-related HG MPNSTs showed the lowest incidence of molecular PRC2 complex abnormalities and exhibited loss of H3K27me3 expression in only 60% of cases, from a practical diagnostic viewpoint this particular clinical subset is rarely in need of a more definitive ancillary marker, as the majority of HG spindle cell sarcomas occurring in an NF1 patient are considered de facto MPNSTs. In addition, in this context most MPNSTs occur nearby or in association with an intraneural or plexiform NF. This scenario is obviously very different in the absence of NF1 clinical history, where the high performance of anti-H3K27me3 IHC could indeed validate the diagnosis of MPNST, otherwise being labeled by default as an unclassified spindle cell sarcoma. Fortunately, the finding of loss of H3K27me3 expression had the highest sensitivity in this particular group with 94% of sporadic and 91% of RT-related MPNSTs. All 5 epithelioid MPNSTs exhibited retention of H3K27me3 expression, further reinforcing that most of these tumors have distinct morphologic, IHC, and molecular features, compared with classic nonepithelioid MPNSTs.

In 50 of 54 (93%) MPNST cases with available IHC and molecular data the H3K27me3 expression status correlated with the EED and/or SUZ12 genetic aberrations. The 4 cases harboring discordant molecular/IHC results were NF1-related MPNSTs. In addition, 1 NF1 patient developing multiple MPNST primaries showed intertumoral heterogeneity at morphologic, H3K27me3 expression and molecular levels. Our study also attempted to investigate the impact of the histologic spectrum of MPNSTs, mostly represented by various degrees of pleomorphism, which typically falls outside the rigidly defined histologic phenotype of MPNST. When focusing on the 17 cases with pleomorphic features, although there was no correlation with the mutational status of the PCR2 components, 70% of these cases showed H3K27me3 loss of expression, particularly the tumors arising in the sporadic (5/6) and RT-related (3/3) clinical settings. Thus, the lack of anti-H3K27me3 staining may help in the diagnosis of MPNSTs with variant morphology, particularly in non-NF1 cases. Most MPNSTs with divergent differentiation demonstrated loss of H3K27me3 expression in the homologous component, with concurrent loss in the divergent component in a subset of cases. However, in 2 MPNSTs with 2 divergent components, whereas the "homologous" portion demonstrated mutations of EED and SUZ12 and concurrent loss of H3K27me3 expression, H3K27me3 positivity was retained in one of the heterologous elements, reinforcing the intratumoral genetic heterogeneity observed in some cases.

To determine the specificity of the H3K27me3 loss of expression in the context of the differential diagnosis of MPNST, we have investigated the pattern of expression of this marker in a large cohort of look-alike lesions. SS and MPNSTs share similar morphologic features, being composed of monomorphic, undifferentiated spindle cells, tightly packed in

intersecting long fascicles. As this morphologic overlap can be quite significant, ancillary methods are often required for a definitive diagnosis. IHC analysis for S100 protein has limited utility in this context, as SS might show patchy reactivity ranging from 15% to 38%. 8–10,26 Although recently 2 additional reliable IHC markers were added in this differential diagnosis—namely, Sox10 and TLE1—their low sensitivity and specificity have perpetuated the diagnostic challenges. Sox10 is positive in only 67% of MPNSTs, while also being present in 7% of SS cases, particularly in intraneural lesions.^{8,27} Similarly, the TLE1 antibody, initially hailed as a sensitive and specific marker for SS, ^{28–30} experienced a subsequent waning of its specificity in later studies,³¹ including focal positivity in up to 30% of MPNSTs.^{29,31} Interestingly, MPNST can potentially express epithelial markers such as focal AE1:AE3 and low-molecular weight keratins, 32 and the MPNST with perineurial differentiation will consistently express epithelial membrane antigen. ^{33,34} The most reliable test in this differential diagnosis has been detecting the t(x;18) translocation in SS, either by reverse transcription polymerase chain reaction of the SS18-SSX1 or SS18-SSX2 gene fusions or by fluorescence in situ hybridization assay for SYT gene rearrangement. 35,36 Nevertheless, molecular testing is not always available, and a prior controversial study has illustrated some of the pitfalls interpreting molecular results, which erroneously suggested that the t(x;18) translocation is not specific for SS but can also occur in MPNST or NFs.³⁷ Our results suggest that H3K27me3 IHC might serve as an additional useful discriminating marker outside the NF1 setting, as all of the SS cases tested (n = 113), including monophasic, biphasic, and poorly differentiated examples, retained expression of this marker. A recent study showed that neurofibromin C-terminus (NFC) antibody, against the protein product of the NF1 gene, is a reliable ancillary marker that differentiates MPNST from other spindle cell neoplasms.³⁸ As the majority of both NF1 and sporadic MPNSTs show NF1 gene mutations, NFC expression is lost in 88% and 43% of cases, respectively. In contrast, all SS cases included in the study (0/22), among several other spindle cell proliferations, showed retention of NFC staining.

The differential diagnosis of MPNST from SCM/DM is also very challenging, especially as these melanoma variants, particularly pure DMs, show no or little expression of melanocytic markers, such as MiTF, Melan A, and HMB45. Although most DM and SCM cases are immunoreactive with S100 protein and Sox10, 8,10,39–41 loss of expression of these markers can occasionally be found, 41,42 particularly in metastatic cases and when the rare phenomenon of dedifferentiation ensues. 43,44 Altered PRC2 components may also play a role in melanoma progression, and they appear to be overwhelmingly composed of activating *EZH2* mutations. 45 Consequentially, a recent study 46 demonstrated an overexpression of H3K27me3, particularly at the tumor front region, in superficial spreading, as well as in nodular, acral, and lentigo maligna melanoma tumors, with no cases of DM/SCM being included in that publication. In this study, all of the DM/SCM melanoma samples tested with H3K27me3 IHC analysis demonstrated retained expression.

Another potential problem is distinguishing deep-seated MPNSTs arising in the trunk or retroperitoneum from a dedifferentiated liposarcoma (DDL), particularly in the 10% to 15% of cases of DDL that will only show an HG spindle cell component. Similarities between DDL and MPNST include a nonspecific immunoprofile, a similar monomorphic spindle cell tumor morphologic picture, and the presence of occasional DDL cases (5% to 10%) with

divergent differentiation, including rhabdomyosarcomatous elements.^{47,48} Nevertheless, although the diagnosis of DDL can be reliably confirmed in most cases by demonstrating either the IHC overexpression or gene amplification of MDM2 and CDK4,⁴⁹ we believe that anti-H3K27me3 IHC could also be helpful in this context. The entirety of liposarcoma samples tested in this study, including 44 examples of dedifferentiated tumors, retained H3K27me3 expression.

Occasionally, GIST might be considered in the differential diagnosis of MPNST, particularly in the NF1 setting, where both GIST and MPNST can occur. Of note, most GISTs in the NF1 setting lack *KIT/PDGFRA* mutations, but they typically express diffuse and strong CD117 reactivity.^{50,51} Rare documented examples of KIT-independent imatinib-resistant GISTs may be seen to lose CD117 expression and show divergent differentiation toward a rhabdomyosarcomatous component.⁵² Our results reveal that none of the GIST samples tested (n = 123), spanning a large clinical and molecular spectrum, showed loss of H3K27me3 expression, adding a potential extra tool when confronted with this differential diagnosis.

Additional spindle cell lesions included in our study were MFS and OFMTs. All of our control cases of MFS and OFMT were immunoreactive with anti-H3K27me3.

In summary, our work advocates the utility of IHC analysis of H3K27me3 as a new and robust diagnostic tool in the armamentarium of soft tissue surgical pathology, in particular in the differential diagnosis of MPNSTs. In certain clinical contexts, such as RT-related or sporadic MPNSTs, loss of H3K27me3 expression emerges as a reliable ancillary marker, with excellent sensitivity and specificity. A lower level of sensitivity of this marker transpires in NF1-related MPNSTs, which is linked to a lower incidence of PRC2 inactivation. None of the entities included in the control groups, which are the most frequent considerations included in the differential diagnosis of MPNSTs, showed H3K27me3 loss of expression. Furthermore, our study indicates that monoclonal antibody clone C36B11 is an appropriate reagent for the analysis of H3K27me3 expression, demonstrating a good correlation between IHC expression and the *EED/SUZ12* mutation status in the vast majority (93%) of cases.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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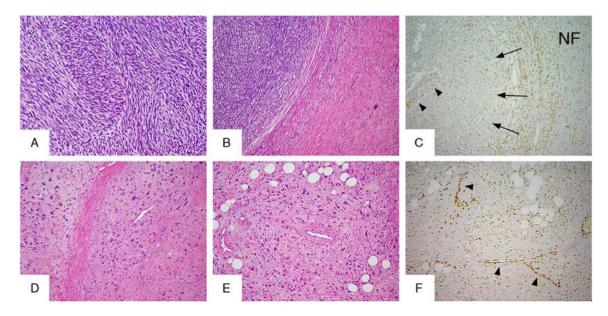


Figure 1.

H3K27me3 immunostaining in morphologic variants of MPNSTs. Classic HG MPNST features with intersecting fascicles of monomorphic spindle cells shown here in an NF1-related tumor (A, B; MPNST03), with an adjacent plexiform NF (B, MPNST03, right side). H3K27me3 immunolabeling showed retained expression in NF (C), whereas the adjacent MPNST exhibited complete loss of expression (C, arrows). Note the internal positive controls: vessels (C, arrowheads), and inflammatory cells retaining staining. An RT-associated HG MPNST (MPNST48.1) with pleomorphic features (D, E) showing loss of H3K27me3 immunostaining (F, note internal control in vessels, arrowheads).

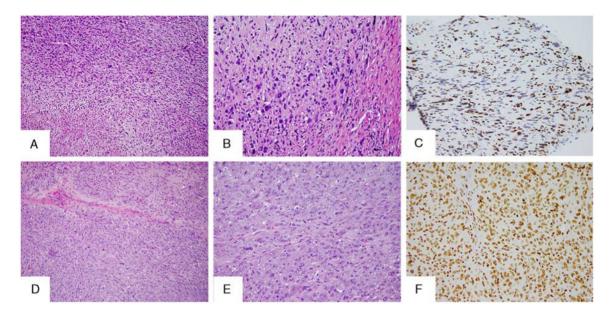


Figure 2. Additional patterns of H3K27me3 immunolabeling in MPNST. Partial loss or mosaic pattern of staining seen in 1 NF1-related tumor (A–C, MPNST12). Note the focal pleomorphism in MPNST12 (A, B). An epithelioid MPNST composed of solid nests of large polygonal cells with eosinophilic cytoplasm and vesicular nuclei with macronucleoli (D, E), showing diffuse immunolabeling (F).

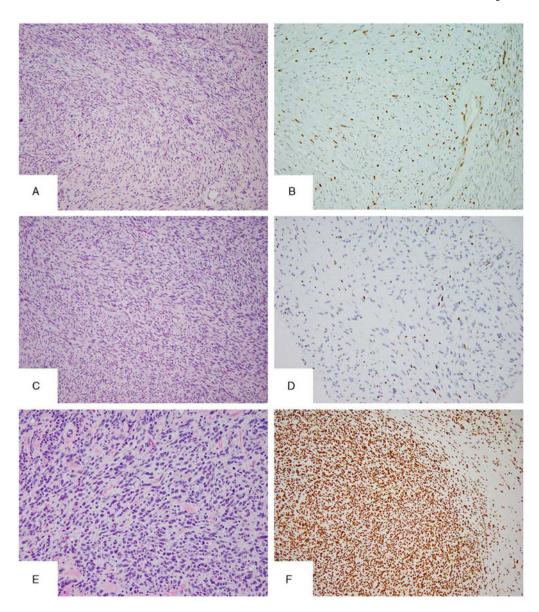


Figure 3. Morphologic and H3K27me3 IHC intertumoral heterogeneity in NF1-related MPNST. This patient (MPNST15) developed 3 different MPNST primaries, which showed variable morphology: HG with focal pleomorphism (MPNST15.1) (A), HG classic (C) arising with an adjacent plexiform NF (MPNST15.3), and HG with pleomorphic features (MPNST15.4) (E). Only the MPNST with pleomorphic features showed retained H3K27me3 (F), whereas the others lost expression (B, D).

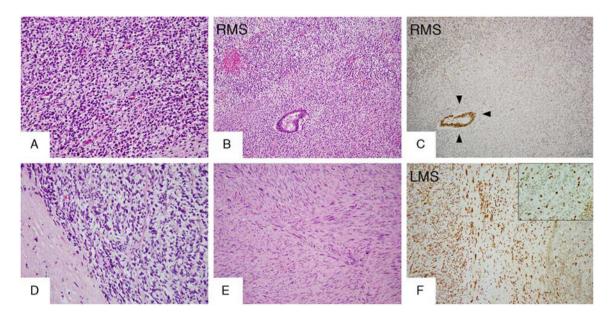


Figure 4. H3K27me3 staining in MPNSTs with heterologous components. A sporadic case (MPNST35) with discordant H3K27me3 expression between the 2 types of divergent differentiation components, rhabdomyosarcomatous (A, B, RMS) and glandular (B): lost expression in the homologous and RMS element (C, RMS) and retained expression in the glandular component (C, arrowheads). H3K27me3 differential expression in another sporadic MPNST (MPNST30) with 2 types of divergent differentiation components: loss of expression in the homologous (small cell) component (D, F right side and inset) and retained expression in both heterologous elements, leiomyosarcoma (E, F left side LMS) and glandular (not shown). LMS indicates leiomyosarcoma; RMS, rhabdomyosarcoma.

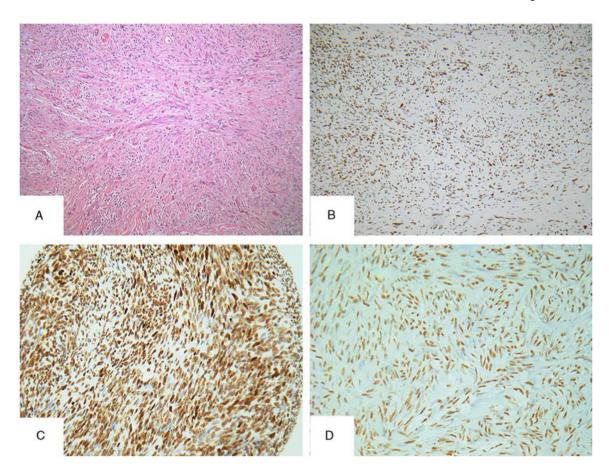


Figure 5. Morphologic mimics of MPNST showed retained H3K27me3 expression. An SCM with long intersecting fascicles of bland monomorphous spindle cells (A) showing diffuse and strong immunoreactivity for H3K27me3 (B). Selected cases of other entities included in the control group with H3K27me3-retained expression: monophasic SS (C) and GIST (D).

Table 1
Summary of the Results of H3K27me3 IHC of the Different MPNST Groups

	H3K27me3 IHC I	Loss (n [%])		
MPNST Groups	Complete	Partial	H3K27me3 IHC Retained	
NF1-associated MPNSTs (N=33)	15 (45)	4 (12)	14 (42)	
Sporadic MPNSTs (N=18)	16 (90)	1 (5)	1 (5)	
RT-associated MPNSTs (N=12)	11 (91)	_	1 (9)	
Epithelioid MPNSTs (N=5)	_	_	5 (100)	
Total MPNST cases (N=68)	47 (69)		21 (31)	

Table 2
H3K27me3 Monoclonal Antibody IHC Results of the Different Entities Included in the
MPNST Differential Diagnosis and Miscellaneous Tumors

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Diagnosis	H3K27me3 IHC Loss/Total Cases		
Cutaneous melanoma			
Pure desmoplastic melanoma	0/37		
Mixed desmoplastic melanoma	0/11		
Spindle cell melanoma	0/5		
Synovial sarcoma (MF, BF, and PD)	0/113		
GIST			
KIT/PDGFRA mutant	0/109		
SDHB-deficient WT pediatric and adult	0/13		
WT dedifferentiated GIST	0/1		
Liposarcoma			
Well differentiated	0/31		
Dedifferentiated	0/44		
Ossifying fibromyxoid tumor	0/6		
Soft tissue myoepithelial carcinomas	0/6		
MFS	0/63		

In the case of GIST, KIT and PDGFRA were WT.

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BF indicates biphasic; MF, monophasic; PD, poorly differentiated.